

## GRAPEFRUIT ALBEDO AND ITS SEED EFFECTS ON DIABETES

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### Abstract

The *Diabetes mellitus* is one of the dominated disease and its focused studies are concerned to compete with this issue worldwide. Grapefruit albedo, seeds and their blends were supplemented in marmalade to evaluate their potential in controlling diabetes and developing immunity. The present study has been planned to find out the grapefruit effect on the *Diabetes mellitus* and immune system. Marmalades supplemented with higher albedo contents i.e. A<sub>5</sub> (containing 5% albedo powder) and A<sub>4</sub>S<sub>1</sub> (containing 4% albedo and 1% seed powder) exhibited higher values for TPCs, TFCs, antioxidant activity as well as satisfactory acceptance by the consumers. These marmalades were further subjected to bio efficacy trials using Sprague Dawley rats induced with diabetes. Rats were divided into 3 groups i.e. G<sub>0</sub> (normal diet + C), G<sub>1</sub> (normal diet + A<sub>5</sub>) and G<sub>2</sub> (normal diet + A<sub>4</sub>S<sub>1</sub>). The G<sub>1</sub> and G<sub>2</sub> exhibited less weight gain with significant decrease in all biomarkers of diabetes including blood glucose level, blood insulin level and HbA1c. Moreover, an increase in red blood cells (16% and 8% in G<sub>1</sub> and G<sub>2</sub>, respectively), hemoglobin (11% and 8.57 %), white blood cells (17% and 6%) and platelet count (1% and 0.7%) was observed. Erythrocyte sedimentation rate and C-reactive protein were decreased by 15% and 5%, respectively, compared to the rats fed with the control diet. These results reflected the potential of grapefruit albedo and seeds in combating diabetes and developing immunity through the development of value-added food products.

**Keywords:** Grapefruit, Marmalade, Sprague Dawley Rats, Diabetes Biomarkers, Management

### Highlights

- Waste materials of grapefruit (albedo and seeds) are ample source of phytochemicals.
- Albedo incorporation improve the organoleptic properties of product.
- Phytochemicals help in curing the diabetes and improve the inflammation status of diabetic patients.

### INTRODUCTION

Pakistan is among the top ten citrus fruit-producing countries in the world. The citrus crop is the leading fruit crop worldwide. Around 124,246 thousand tonnes of citrus were produced in 2016 (FAO, 2016). China, Brazil, Spain, Mexico, India and the United States are major citrus-producing countries that contribute almost two third of global citrus

production. Among citrus fruits, mandarins, lemon, oranges, mosambi and grapefruit are widely consumed (Liu, Heying, & Tanumihardjo, 2012). Grapefruit (*Citrus paradisi*) is the oldest citrus fruit known for its sweet and somewhat sour taste. It is grown mostly in tropical and subtropical areas. Grapefruits have several promoting benefits due to bioactive phytochemicals like organic acids, furanocoumarins, coumarins, carotenoids, flavonoids and polyphenols. A grapefruit has more than 188 mg potassium, which can fulfill the body's daily requirements and provides sufficient Vitamin C to improve the immune system's status. The high level of pectin and fiber found in citrus fruits may help to maintain healthy cholesterol and glucose levels (Faid, Fadlalla, & Khojah, 2017). White inner layer (albedo) and outer colored layer (flavedo) of grapefruit peels are rich in soluble and insoluble dietary fibers, which can be applied in the food industry for making the value added nutraceutical products (Khalil, Farghal, & Farag, 2020). On average, 34-50% of citrus fruits are processed into juices which produce large amounts of peels and seeds as waste (around 40 to 50% weight of fresh fruit), mostly used as cattle feed (Chavan, Singh, & Kaur, 2018; Zarate-Vilet, Gué, Delalonde, & Wisniewski, 2022).

Grapefruit peels are an ample source of potential nutraceutical components, including flavonoids (catechin, epigallocatechin, rutin, kampferol, flavonone, flavanone glycosides, polymethoxylated flavones, naringin, naringenin, limonin and quercetin) and phenolics (gallic acid, caffeic acid, ferulic acid, sinapinic acid, hesperidin and neohesperidin). These bioactive components vary from 0.5-20% dry basis in peels. Bioactive components in fresh and dry albedo exhibit pharmacological benefits such as anti-inflammatory, antimicrobial, antibacterial, anti-cancerous, and anti-oxidant and protect the body from multiple organ injuries (Ademosun et al., 2015; Scurria et al., 2021). These also inhibit prostate cancer and reduce the side effects of drugs used during oesophageal carcinoma chemotherapy (Nishad et al., 2020). Grapefruit is also a rich source of bioactive compounds such as vicenin-2, naringenin, rhoifolin, neohesperidin, hesperidin and didymin (Avula et al., 2016).

Almost 7.1 million people in Pakistan are suffering from diabetes, which is increasing daily. Around 46-75% Pakistani diabetic people have metabolic syndrome. Diabetic people are more prone to infections. The course of infection is also more complicated in diabetic patients. High blood sugar levels can weaken the immune system. Furthermore, some microorganisms become more virulent in high-glucose environments (Sarfraz, Sajid, & Ashraf, 2016). Secondary metabolites and bioactive components of grapefruit possess antidiabetic effects. Soluble and insoluble dietary fibers in grapefruit peels and seeds act as prebiotics and enhance the gut microbial activity that helps improve immunity status (Vazquez-Olivo, Gutiérrez-Grijalva, & Heredia, 2019).

Previously, the raw peel/seed and their extracts have been studied for their application against metabolic diseases, including oxidation stress (Huang, Liao, Qi, Jiang, & Yang, 2020; Sir Elkhatim, Elagib, & Hassan, 2018), obesity (Farouk, El-Sayeh, Mahmoud, & Sharaf, 2012), hyperchlostermia (Fayek et al., 2017), liver failure (Udom et al., 2018) and diabetes (Gan et al., 2020; Oyetayo, Akomolafe, & Oladapo, 2019). The current study

comprises of development of grapefruit albedo and seed-based functional marmalade and its effects on diabetes and immunity.

## MATERIALS AND METHODS

Grapefruits (pink variety) were procured from the local market of Faisalabad. The grapefruits were selected randomly based on certain quality attributes, including color uniformity, being free from abrasion, size and shape. For biological efficacy analysis, diagnostic kits were procured from Sigma-Aldrich bioassay and cayman chemicals (Cayman Europe, Estonia). All other chemicals and reagents were of analytical grade.

**Preparation of albedo and seed powder:** Grapefruits were peeled manually with the help of a knife. Peels, seeds and pulp were separated and kept in different pots. Albedo (white portion of peels) was separated from Flavedo. The albedo and seeds were dried in Hot Air Oven at 65 °C for 48 hours. After drying, they were subjected to milling by a rotor mill (ZM 200, Retsch GmbH, Haan, Germany). The resulting powders were packed in polyethylene bags for further use.

**Preparation of marmalade:** Grapefruit marmalade was prepared according to the method described by Tamer (2012) with some modifications. Different concentrations of albedo and seed powders were incorporated in marmalades, as given in table 1.

**Table 1: Various concentrations of albedo and seed powders added to product**

Treatment designation	Description
C	Control
A <sub>5</sub>	Albedo powder (5%)
S <sub>5</sub>	Seed powder (5%)
A <sub>2.5</sub> S <sub>2.5</sub>	Albedo powder (2.5%), Seed powder (2.5%)
A <sub>1</sub> S <sub>4</sub>	Albedo powder (1%), Seed powder (4%)
A <sub>4</sub> S <sub>1</sub>	Albedo powder (4%), Seed powder (1%)

### **Characterization of marmalade:**

Flavonoid contents, phenolic contents (Folin-Ciocalteu method) and antioxidant activity (DPPH method) of all marmalade samples were determined by the methods described by Chang et al. (2007), Sun et al. (2007) and Sreeramulu, Reddy, and Raghunath (2009), respectively described. The sensory quality of marmalades was evaluated by using the 9-point hedonic scale (9=like extremely, 1=dislike extremely) according to the method described by Lawless and Heymann (2010).

**Bio-Efficacy trials:** The treatments exhibiting better phytochemical and sensory characteristics were further subjected to *in vivo* trials using animal models. Bio-efficacy trials were performed using Sprague Dawley rats to explore the anti-hyperglycemic potential and functional worth of selected treatments. Purposely, experimental rats were acclimatized individually under optimum conditions (23±1 °C temperature and relative humidity 55-60%) in stainless steel cages. All animals were fed standard diet and tap water *ad libitum* (physiological saline) for seven days (one week) before experimentation.

**Study plan on hyperglycemic rats:** Rats (n=18) were randomly divided into 3 groups (G<sub>0</sub>, G<sub>1</sub> and G<sub>2</sub>). Streptozotocin injection (60 mg/kg) was administered intraperitoneally to induce diabetes in rats. G<sub>0</sub> was administered with control marmalade (C) whereas, G<sub>1</sub> and G<sub>2</sub> were fed on A<sub>5</sub> and A<sub>4</sub>S<sub>1</sub> respectively, incorporated with a normal diet (Table 1). After 30 days, overnight fasted rats were weighed, slaughtered and their sera were collected to assess their diabetic and infection status.

**Calculations of feed and drink intake:** During the entire study period, rats' feed and drink intake were calculated daily as described by Wolf and Weisbrode (2003) and Slemmer, Shaughnessy, Scanlan, Sweeney, and Gottschall-Pass (2012). The body weight and net feed intake were used to determine the feed efficiency ratio using the given mathematical relation.

$$FER \text{ (Feed Efficiency Ratio)} = \frac{\text{Body weight Gain (g/day)}}{\text{Feed intake (g/day)}} \times 100$$

**Biomarkers for Diabetes mellitus:** Fasting blood glucose level (mg/dL) was determined through compatible strips employing the commercially available glucometer (OnCall® Ez II; SN 303S0014E09) (DeMambro et al., 2010; Dong, Cai, Shen, & Liu, 2011). Blood insulin level was measured by Enzyme-Linked Immunosorbent Assay (ELISA Kit R&D system) (Ahn, Choi, Kim, & Ha, 2011). HbA1c levels were checked at baseline and the end of the study (30 days) (Slemmer et al., 2012).

**Biomarkers for inflammation response evaluation:** Hematological assay *i.e.* erythrocytes, leukocyte and thrombocyte indices were analyzed using Medonic M Series hematology analyzer (Boule Diagnostics Int. AB Stockholm, Sweden). Moreover, Erythrocyte Sedimentation Rate was assessed using Westergren's method using an automated ESR System (Gerard et al., 2015; Karbiner et al., 2013; Plebani & Piva, 2002; Uzun & Kalender, 2013) C-reactive protein was determined through the immunoturbidimetry method according to the procedure described in Putra, Saputro, Nurrahman, Herawati, and Dewi (2020).

**Statistical analysis:** A complete randomized design (2-factor factorial) was applied for animal study parameters and a simple complete randomized design was used for product characterization parameters using Statistix 8.1 model. Furthermore, Tukey's HSD multiple comparison test was applied for means comparison following the principles outlined by Montgomery (2017).

## RESULTS AND DISCUSSION

### Results

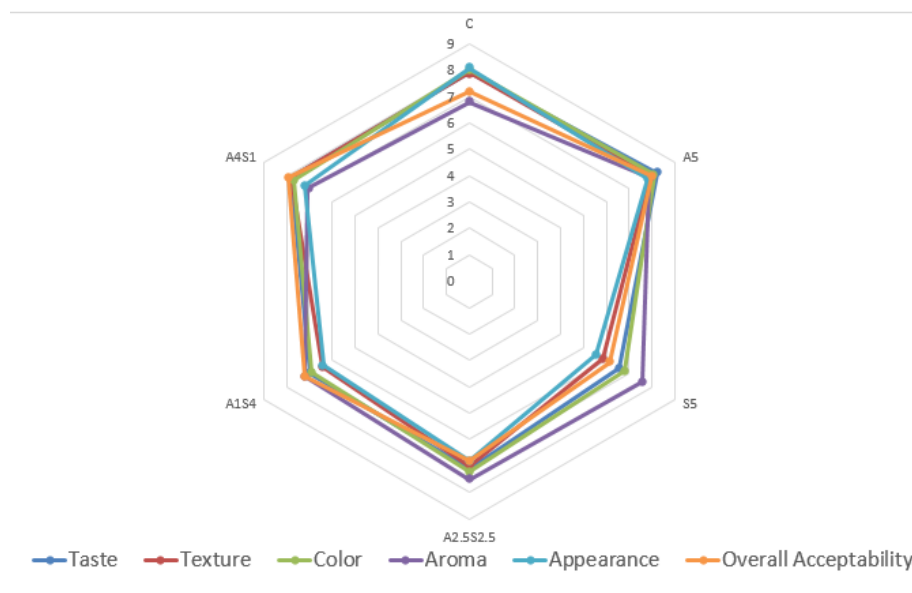
**TFCs, TPCs and DPPH assay of marmalades:** The mean values of total flavonoids, total phenols, and antioxidant activity of supplemented marmalades have been summarized in Table 2. The results show that the addition of albedo and seeds significantly affected the TPCs, TFCs and DPPH antioxidant activity of marmalades ( $p < 0.05$ ). All supplemented marmalades showed significant differences in mean values

of TPCs and DPPH antioxidant activity. The minimum TPCs were observed in the control marmalade (121.3 mg GAE/100g) while maximum TPCs were observed in A<sub>5</sub> marmalade (222.6 mg GAE/100g). Similarly, minimum TFCs were observed in C (0.46 mg QE/100g) and maximum TFCs were recorded in A<sub>5</sub> (1.9 mg QE/100g). The antioxidant activity was decreased by decreasing the contents of albedo powder in treatments. Maximum antioxidant activity was exhibited by A<sub>5</sub> followed by A<sub>4</sub>S<sub>1</sub>, A<sub>2.5</sub>S<sub>2.5</sub>, A<sub>1</sub>S<sub>4</sub>, S<sub>5</sub> and C. Among all supplemented marmalades, samples containing higher albedo concentration had higher TPCs, TFCs and DPPH antioxidant (%) activity compared to marmalades having more seed proportion.

**Table 2: Mean values of TFCs, TPCs and DPPH assay of supplemented marmalades with albedo and seeds**

Phytochemistry	Treatments					
	C	A <sub>5</sub>	S <sub>5</sub>	A <sub>2.5</sub> S <sub>2.5</sub>	A <sub>1</sub> S <sub>4</sub>	A <sub>4</sub> S <sub>1</sub>
TPC (mg GAE/100g)	121.3±0.5 <sup>f</sup>	222.6±2.7 <sup>a</sup>	168.7±3.3 <sup>e</sup>	186.4±2.2 <sup>c</sup>	179.4±1.4 <sup>d</sup>	201.3±1.4 <sup>b</sup>
TFC (mg QE/100g)	0.46±0.02 <sup>d</sup>	1.9±0.02 <sup>a</sup>	0.98±0.1 <sup>c</sup>	1.4±0.04 <sup>b</sup>	1.37±0.2 <sup>b</sup>	1.59±0.08 <sup>b</sup>
DPPH (%)	12.13±0.3 <sup>f</sup>	24.8±0.4 <sup>a</sup>	14.6±0.29 <sup>e</sup>	18.8±0.3 <sup>c</sup>	16.8±0.3 <sup>d</sup>	20.3±0.7 <sup>b</sup>

**Sensory evaluation:** The addition of albedo and seed powders in marmalade significantly influenced the taste, color and texture. Maximum scores for taste, texture, aroma, appearance, overall acceptability and color were assigned to A<sub>5</sub> and A<sub>4</sub>S<sub>1</sub>. While other supplemented marmalades attained lower organoleptic scores. The sensory scores of organoleptic properties of all marmalades have been graphically represented in Figure 1.



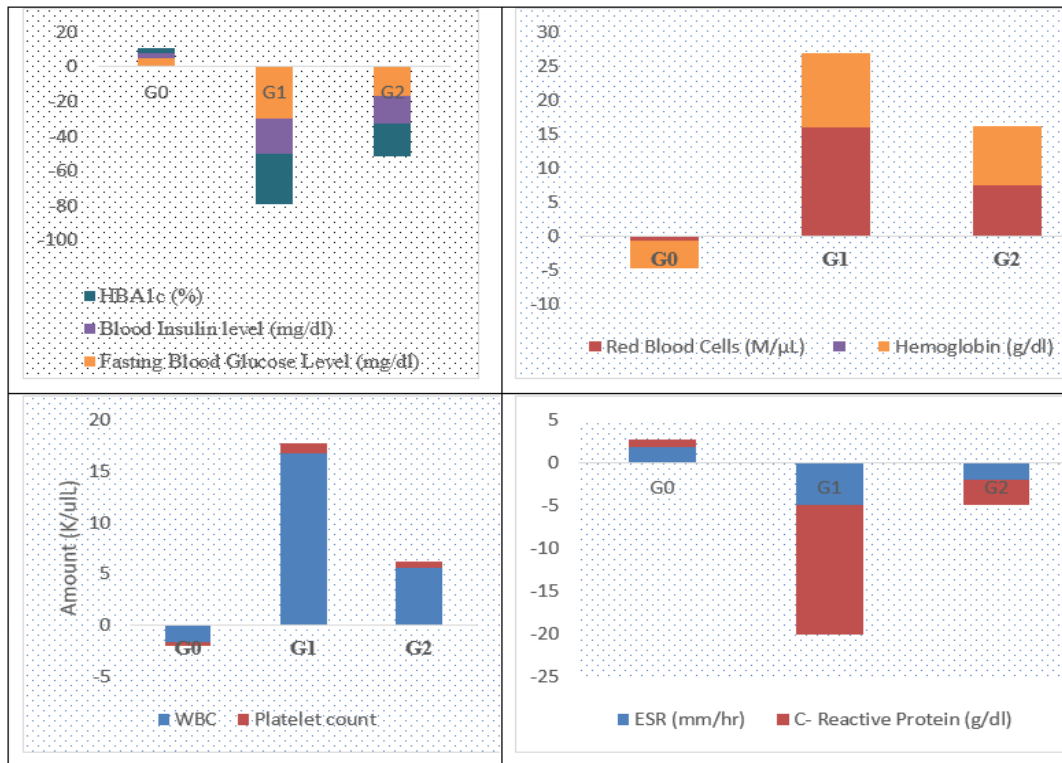
**Figure 1: Sensory evaluation of supplemented marmalades with grapefruit albedo and seeds**

**Calculations of feed and drink intake:** The average weight gain, feed intake and water intake were higher in the control group compared to the treated groups (G<sub>1</sub> and G<sub>2</sub>), as shown in Table 3. These symptoms are commonly known markers of type 2 diabetes due to insulin deficiency in diabetic rats. The average feed intake, water intake and weight gain were significantly reduced in rats treated with albedo and seed powders supplement. The average weight gain (g/day) in experimental groups *i.e.* G<sub>0</sub>, G<sub>1</sub> and G<sub>2</sub> was 0.24, 0.112 and 0.208 (g/day), respectively. The rats fed with A5 showed lower feed intake, water intake and weight gain values compared to A4S1.

**Table 3: Effect of supplemented marmalades on growth performance parameters**

Experimental Groups	Growth Performance Parameters					
	Initial Body Weight (g)	Final Body Weight (g)	Weight gain (g/day)	Average Feed Intake (g/day)	Feed Efficiency Ratio (FER)	Average Water Intake (mL/day)
G <sub>0</sub>	137.57±8.8 <sup>c</sup>	140.54±5.2 <sup>c</sup>	0.24±0.013 <sup>a</sup>	19.14±0.58 <sup>a</sup>	1.24±0.65 <sup>a</sup>	24.27±0.47 <sup>a</sup>
G <sub>1</sub>	153.41±2.9 <sup>a</sup>	156.85±2.8 <sup>a</sup>	0.112±0.01 <sup>c</sup>	16.44±0.37 <sup>c</sup>	0.68±0.07 <sup>a</sup>	21.16±0.76 <sup>c</sup>
G <sub>2</sub>	142.64±3.0 <sup>b</sup>	149.04±3.2 <sup>b</sup>	0.208±0.02 <sup>b</sup>	17.72±0.61 <sup>b</sup>	1.17±0.12 <sup>a</sup>	23.33±0.59 <sup>b</sup>

Values are expressed as mean±SD; one-way ANOVA followed by Tukey's HSD test ( $p \leq 0.05$ ).



**Figure 2: Graphical representation of effect of supplemented marmalades with albedo and seeds on biomarkers for *Diabetes mellitus* and immunity**

**Biomarkers for Diabetes mellitus:** It is evident from the results (shown in Table 4) that rats of G<sub>0</sub> had maximum fasting blood glucose level (261.6 mg/dl) followed by G<sub>2</sub> (194.8 mg/dl) and G<sub>1</sub> (182.8 mg/dl). Higher blood insulin level was observed in G<sub>0</sub> (35.98 units/l) followed by G<sub>2</sub> (26.5 units/l) and G<sub>1</sub> (23.14 units/l). Similarly, HbA1c was higher in G<sub>0</sub> (6.89 %) trailed by G<sub>2</sub> (4.68 %) and G<sub>1</sub> (4.39 %) at the end of the trial. The graphical representation of the results can be seen in figure 2. Fasting blood glucose level, blood insulin level and HbA1c were higher in the control group (G<sub>0</sub>). In comparison, a significant decrease was observed in all biomarkers of *Diabetes mellitus* in G<sub>1</sub> and G<sub>2</sub> groups. These results indicate that the addition of grapefruit albedo and seeds in marmalade played a significant role in controlling diabetes. Moreover, the marmalades supplemented with albedo showed more antidiabetic potential than marmalades supplemented with seeds.

**Table 4: Mean values of biomarkers for *Diabetes mellitus***

	Fasting Blood Glucose Level (mg/dl)		Biomarkers for <i>Diabetes mellitus</i>						
	Day (0)	Day (30)	% Change	Blood Insulin Level (units/l)			HbA1c (%)		
				Day (0)	Day (30)	% Change	Day (0)	Day (30)	% Change
<b>G<sub>0</sub></b>	249.54± 4.11 <sup>b</sup>	261.6±6.8 <sup>a</sup>	+4.83	34.9± 2.9 <sup>a</sup>	35.98± 1.16 <sup>a</sup>	+3.09	6.7± 0.4 <sup>a</sup>	6.89± 0.31 <sup>a</sup>	+2.8
<b>G<sub>1</sub></b>	260.43± 1.24 <sup>a</sup>	182.8±5.4 <sup>e</sup>	-30	28.9± 0.8 <sup>c</sup>	23.14± 1.25 <sup>e</sup>	-20	6.2± 0.14 <sup>b</sup>	4.39± 0.05 <sup>f</sup>	-29
<b>G<sub>2</sub></b>	233.7± 10.54 <sup>c</sup>	194.8±3.9 <sup>d</sup>	-17	31.6± 1.14 <sup>b</sup>	26.5± 0.97 <sup>d</sup>	-16	5.8± 0.1 <sup>c</sup>	4.68± 0.1 <sup>e</sup>	-19

Values are expressed as mean±SD; Two-way ANOVA followed by Tukey's HSD test ( $p \leq 0.05$ )

**G<sub>0</sub>** Normal diet+C

**G<sub>1</sub>** Normal diet+A<sub>5</sub>

**G<sub>2</sub>** Normal diet+A<sub>4</sub>S<sub>1</sub>

**C** = Control; **A<sub>5</sub>** = Grapefruit Marmalade containing 5% albedo powder; **A<sub>4</sub>S<sub>1</sub>**= Grapefruit Marmalade containing 4% albedo and 1% seeds powder

**-ve**= Improvement      **+ve**= No Improvement

**Biomarkers for immune response evaluation:** Effect of supplemented marmalades on erythrocyte indices, thrombocyte count, leukocyte count, inflammatory biomarkers, Erythrocyte Sedimentation Rate (ESR) and C-reactive protein have been represented in Tables 5, 6 and 7 respectively. The graphical representation of the results can be seen in figure 2. It is evident from the results that treatments significantly affected the red blood cells and hemoglobin in diabetic rats ( $p < 0.05$ ). Likewise, treatments significantly affected the white blood cells, platelet count and C-reactive protein ( $p < 0.05$ ). However, the effects of treatments on erythrocyte sedimentation rate in diabetic rats was non-significant. It was also observed that the combined effect of treatments and days significantly influenced the red blood cells, hemoglobin and white blood cells. Whereas, treatments and days

interaction was non-significant in platelet count, erythrocyte sedimentation rate and C-reactive protein.

During the experiment, a maximum increase in total red blood cells was observed in G<sub>1</sub> (+16%) trailed by G<sub>2</sub> (+8%). However, a decrease in red blood cells was recorded in G<sub>0</sub> (-0.6%). Likewise, the highest increase in hemoglobin was observed in G<sub>1</sub> (+11%) followed by G<sub>2</sub> (+8.57%). However, a decrease in hemoglobin was recorded in G<sub>0</sub> (-4%). Similarly, a maximum increase in thrombocyte indices *i.e.* white blood cells was observed in G<sub>1</sub> (+17%) followed by G<sub>2</sub> (+6%); whereas, decrease in white blood cells was noted in G<sub>0</sub> (-2%). Control group showed decrease in platelet count (-0.37%). However, an increase in platelet count was observed in G<sub>1</sub> (+1%) and G<sub>2</sub> (564.68 to 568.79 K/ $\mu$ L; +0.7%).

The mean values of inflammatory biomarkers at base line and end of experiments showed that inflammation status in the control group (G<sub>0</sub>) was increased. The erythrocyte sedimentation rate in G<sub>0</sub> was increased (+1.83%). G<sub>1</sub> and G<sub>2</sub> groups showed a significant decrease in erythrocyte sedimentation rate. A higher decrease in erythrocyte sedimentation rate was observed in G<sub>1</sub> (-5%) followed by G<sub>2</sub> (-2%). Similarly, the C-reactive protein level was increased in the control group (+0.91%). Whereas, the level of C-reactive protein was decreased in G<sub>1</sub> (-15%) and G<sub>2</sub> (-3%).

**Table 5: Effect of marmalade diet on erythrocyte indices of experimental rats**

Experimental groups	Erythrocyte indices					
	Red Blood cells (M/ $\mu$ L)			Hemoglobin (g/dl)		
	Day (0)	Day (30)	% Change	Day (0)	Day (30)	% Change
G <sub>0</sub>	6.44 $\pm$ 0.29 <sup>de</sup>	6.40 $\pm$ 0.9 <sup>e</sup>	-0.6	11.5 $\pm$ 0.67 <sup>d</sup>	11.05 $\pm$ 1.5 <sup>d</sup>	-4
G <sub>1</sub>	6.96 $\pm$ 0.11 <sup>c</sup>	8.28 $\pm$ 0.31 <sup>a</sup>	+16	12.54 $\pm$ 0.39 <sup>bc</sup>	14.09 $\pm$ 0.2 <sup>a</sup>	+11
G <sub>2</sub>	6.87 $\pm$ 0.2 <sup>cd</sup>	7.43 $\pm$ 0.29 <sup>b</sup>	+8	11.85 $\pm$ 0.62 <sup>cd</sup>	12.95 $\pm$ 0.3 <sup>b</sup>	+8.57

**Table 6: Effect of supplemented marmalade on WBCs and thrombocyte count**

Experimental groups	White Blood cells (K/ $\mu$ L)			Platelet count (K/ $\mu$ L)		
	Day (0)	Day (30)	% Change	Day (0)	Day (30)	% Change
G <sub>0</sub>	8.12 $\pm$ 0.43 <sup>b</sup>	7.99 $\pm$ 1.12 <sup>b</sup>	-2	548.51 $\pm$ 11.3 <sup>b</sup>	546.47 $\pm$ 14.61 <sup>b</sup>	-0.37
G <sub>1</sub>	8.5 $\pm$ 0.2 <sup>b</sup>	10.2 $\pm$ 0.5 <sup>a</sup>	+17	567.78 $\pm$ 3.55 <sup>a</sup>	573.58 $\pm$ 3.98 <sup>a</sup>	+1
G <sub>2</sub>	8.17 $\pm$ 10.54 <sup>b</sup>	8.65 $\pm$ 0.66 <sup>b</sup>	+6	564.68 $\pm$ 10.3 <sup>a</sup>	568.79 $\pm$ 10.6 <sup>a</sup>	+0.7



**Table 7: Effect of supplemented marmalades with albedo and seeds on inflammatory biomarkers**

Experimental groups	Inflammatory Biomarkers					
	ESR (Erythrocyte Sedimentation Rate) mm/hr			C-reactive Protein (g/dl)		
	Day (0)	Day (30)	% Change	Day (0)	Day (30)	% Change
<b>G<sub>0</sub></b>	5.696±0.25 <sup>b</sup>	5.8±0.32 <sup>a</sup>	+1.83	7.68±0.42 <sup>a</sup>	7.75±0.8 <sup>ab</sup>	+0.91
<b>G<sub>1</sub></b>	5.723±0.15 <sup>a</sup>	5.42±0.29 <sup>e</sup>	-5	7.39±0.4 <sup>c</sup>	6.26±0.37 <sup>e</sup>	-15
<b>G<sub>2</sub></b>	5.729±0.14 <sup>c</sup>	5.60±0.11 <sup>d</sup>	-2	7.6±0.7 <sup>b</sup>	7.4±0.65 <sup>d</sup>	-3

Values are expressed as mean±SD; Two-way ANOVA followed by Tukey's HSD test ( $p \leq 0.05$ )

**G<sub>0</sub>** Normal diet+C

**G<sub>1</sub>** Normal diet+A<sub>5</sub>

**G<sub>2</sub>** Normal diet+A<sub>4</sub>S<sub>1</sub>

**C** = Control; **A<sub>5</sub>** = Grapefruit Marmalade containing 5% albedo powder; **A<sub>4</sub>S<sub>1</sub>**= Grapefruit Marmalade containing 4% albedo and 1% seeds powder

**-ve**= Decreasing (Improvement) **+ve**= Increasing (No improvement)

## DISCUSSION

The mean values of total flavonoids, total phenols, and antioxidant activity of supplemented marmalades have been summarized in Table 2. The results show that the addition of albedo and seeds significantly affected the TPCs, TFCs and DPPH antioxidant activity of marmalades ( $p < 0.05$ ). All supplemented marmalades showed significant differences in mean values of TPCs and DPPH antioxidant activity. Estaji, Mohammadi-Moghaddam, Gholizade-Eshan, Firoozzare, and Hooshmand-Dalir (2020) studied the effect of plum peel on the bioactivity of plum marmalade. Authors reported that phenolic compounds (38.65-212.69 mg GAE/100g) and antioxidant activity (33.65-85.47%) in plum marmalade were improved as the concentration of plum peels was increased. Additionally, Ben-Rejeb, Dhen, Kassebi, and Gargouri (2020) prepared the grapefruit jelly and reported that grapefruit jelly had a significant amount of total phenols (123.16 mg GAE/100g), total flavonoids (1.54 mg QE/100g) and exhibited high antioxidant activity. Another study conducted research on watermelon candies fortified with different concentrations of orange albedo and flavedo. It was concluded that fortified candy formulations had higher TPC, TFC and antioxidant activity (Marinelli et al., 2020).

Among all supplemented marmalades, samples containing higher albedo concentration had higher TPCs, TFCs and DPPH antioxidant (%) activity compared to marmalades having more seed proportion. This observation could be due to the presence of more TPCs, TFCs and DPPH antioxidant (%) activity in grapefruit albedo as compared to seeds, as also reported by Assefa and Keum (2017); Sir Elkhatim et al. (2018) and Wang et al. (2020).

Sensory response significantly influences product demand, consumer acceptability and purchase intent. Quality of food products is examined by complex sensation resulting from the interaction of human senses *i.e.* smell, touch, taste, etc. (Ben-Rejeb et al., 2020). The addition of albedo and seed powders in marmalade significantly influenced the taste, color and texture. The more concentration of seeds powder reduced the appearance, aroma, color, taste and overall acceptance of marmalade. This could be due to compounds in seeds powder, including poncirin, hesperidin, polymethoxylated flavones and non-hesperidin which are mainly responsible for the taste change (Kiefl et al., 2017).

Similar results were reported by Teixeira et al. (2020) in which orange jam containing 4% orange peels was reported to be highly acceptable by consumers as compared to other treatments containing 8 and 12% peels proportion. In another study, Younis, Islam, Jahan, Yousuf, and Ray (2015) prepared the papaya jams by incorporating the 2.5, 5, 7.5, 10 and 12.5% mosambi peels. The authors reported that papaya jam having 5% mosambi peels was more acceptable by consumers. Present results are also by the findings of Levent (2014) who incorporated 5% lemon, orange and grapefruit albedo in tarhana soup and reported acceptable sensory response. Other reports on sensory response similar to present results can be found in Marinelli et al. (2020), Özbek, Şahin-Yeşilçubuk, and Demirel (2019), Pérez-Herrera, Martínez-Gutiérrez, León-Martínez, and Sánchez-Medina (2020) and Hussein, Kamil, Hegazy, Mahmoud, and Ibrahim (2015). These symptoms are commonly known markers of type 2 diabetes due to insulin deficiency in diabetic rats. This effect could be due to the higher amount of fibers in the albedo as compared to seeds (Teixeira et al., 2020).

Zhang et al. (2020) described that the anti-obesity potential of citrus peels could be due to dynamic changes in gut microbiota and the improvement of biological processes of metabolism. It significantly increases the amount of fecal short-chain fatty acids *i.e.* acetic acid (43%) and propionic acid (86%). It also decreases the prevalence of proteobacteria and the amount of firmicutes to bacteroidetes ratio, which significantly affects the obesity pathway. These results were similar to the findings of Oyedemi, Adewusi, Aiyegoro, and Akinpelu (2011) who elucidated the antidiabetic effect of *Azelia Africana* (mahogany plant) in controlling the desire for water and feed intake under diabetic conditions. Similar findings were reported by Kang, Song, Lee, Chang, and Lee (2018). The authors elucidated the significant decrease in weight, BMI, triglycerides, cholesterol and low-density lipid profile in 118 patients taking citrus (*Citrus unshiu*) peel pellets for 4 weeks.

Type 2 diabetes is a complicated metabolic disorder denoted by glucose intolerance and hyperglycemia. Increasing evidence from animal models and cells suggests that bioactive components present in peels and seeds may directly ameliorate hyperglycemia, preventing amyloid aggregation and enhancing the insulin secretion by pancreatic  $\beta$  cells (Li, Zhang, Rasool, Geetha, & Babu, 2019). The present study's results corroborate the findings of Oboh, Olasehinde, and Ademosun (2017). The authors expounded the antidiabetic effect of essential oils extracted from peels of orange and lemon. The results revealed the significant inhibitory effect of essential oils on  $\alpha$ -amylase and  $\alpha$ -glucosidase. Comparable findings were also documented by Ali et al. (2020). The authors elucidated

the antidiabetic, anti-hyperlipidemia and antioxidant potency of fruit peels of *Citrus reticulata* in nicotinamide /streptozotocin-induced type 2 diabetes Wistar rats. According to Gan et al. (2020), *in-vitro* digestive glucose rate of bread incorporated with soluble dietary fibers of grapefruit peels was decreased significantly. In another study, Ashraf, Butt, Iqbal, and Suleria (2017) investigated the hypoglycemic potential of citrus (orange) peel powder through rodent modeling. It was concluded that the nutraceutical diet alleviated serum glucose by 8.96% and enhanced insulin secretion by 5.41% in rodents. In another study, serum glucose level was decreased significantly in fish by feeding them *Citrus bergamia* peel oil along with a normal diet (Kesbiç, Acar, Yilmaz, & Aydin, 2020). The mechanism of grapefruit waste in controlling diabetes includes the upregulation of insulin secretion capability as well as intracellular insulin concentration in RIN-m5F cells and Glut4 in HepG2 cells (Lin, Huang, Chen, & Peng, 2021). Pectin obtained from grapefruit peel and other citrus waste showed inhibition of enzymes (against  $\alpha$ -glucosidase, pancreatic cholesterol esterase and lipase) linked to managing hyperglycemia and hyperlipidemia and hypercholesterolemia.  $\alpha$ -glucosidase is responsible for the hydrolysis of glucose in the body, which prevents the postprandial glucose peaks from playing a key role in the management of type- 2 diabetes (Picot-Allain et al., 2022).

It is evident from the results that treatments significantly affected the red blood cells and hemoglobin in diabetic rats ( $p < 0.05$ ). Whereas, treatments and days interaction was non-significant in platelet count, erythrocyte sedimentation rate and C-reactive protein. The present findings are in harmony with the findings of Tsujiyama, Mubassara, Aoshima, and Hossain (2013) who reported that aqueous peel extract of grapefruit suppressed the inflammation in mice cavities. The peel extract (0.1 mL/10g) was administered orally in mice. The anti-inflammatory action of grapefruit peel extract was comparable to indomethacin (anti-inflammatory drug) in mice. Hence, grapefruit peel extract could be utilized to develop anti-inflammatory supplements and foods. Another study investigated the chemical and pharmacological potential of citrus peel oils through *in vitro* and *in vivo* analysis. It was revealed that Citrus limetta peels inhibited the production of pro-inflammatory cytokines, lipid peroxidation, C-reactive protein, ameliorate the histological damage and reactive oxygen species production in white rabbits and mice (experimental animals). *In vitro* and *in vivo* analysis of citrus peels showed that they are safe for topical application on the skin (Maurya, Mohanty, Pal, Chanotiya, & Bawankule, 2018). The current study's results align with earlier investigations of Jiang et al. (2019), who studied the anti-inflammatory and hepatoprotective effects of citrus peels in rats. It was reported that citrus peel extract suppressed the activities of phosphorylated cytokines (by inhibiting the systemic and intrahepatic inflammation), significantly attenuated the hepatic lesions and decreased the non-alcoholic fatty liver disease activity scores. It was documented by Ashraf et al. (2017) that red blood cells, white blood cells and platelet count in rodents were increased by the intake of orange peel powder and orange peel extract with time.

Adding albedo and seeds improved the hematological parameters (red blood cells, hemoglobin, platelet count and white blood cells). This effect was recorded due to the presence of phenolic compounds (polyphenols, hydrocarbons, terpenes), which could

inhibit the hemolysis of red blood cells and lipid peroxidation of cells in diabetic rats (Soji-Omoniwa, Muhammad, Omoniwa, & Usman, 2014). Flavonoids (hesperidin and naringin) present in albedo and seeds stimulate erythropoietin secretion, enhancing the production of red blood cells (Mahmoud, 2013). The platelet count in diabetic-treated rats was improved markedly. This observation indicated albedo and seeds' ability to stimulate the clotting factors' synthesis (Oyedemi et al., 2011). Thus, the current study revealed that albedo and seeds could prevent -associated diabetes inflammation by attenuating the hematological parameters, erythrocyte sedimentation rate and C - reactive protein levels in diabetic rats.

## CONCLUSION

Using underutilized parts of fruits in foods can reduce the negative impact of organic waste disposal on the environment and promote the consumer access to healthier foods. The present study evaluated the nutritional benefits of grapefruit albedo and seeds in controlling *Diabetes mellitus* and inflammation. The study concluded that albedo has more phenolic contents, flavonoids and antioxidants than seeds. Outcomes of sensory analysis and efficacy study proved that the use of albedo powder supplemented marmalade (A5) and seeds (A4S1) is efficient against diabetes as well as inflammation. However, the addition of 5% seeds powder reduced the consumer acceptability of marmalade as well as proved less effective against diabetes as compared to albedo. Supplemented marmalade effectively controlled diabetes by improving the hematological and inflammation parameters.

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## Conflict of Interest

All authors declare that there are no conflict of interest.

## Ethical statement

The ethical committee of the University of Agriculture Faisalabad-Pakistan approved the study.

## References

- 1) Ademosun, A. O., Oboh, G., Passamonti, S., Tramer, F., Ziberna, L., Boligon, A. A., & Athayde, M. L. (2015). Phenolics from grapefruit peels inhibit HMG-CoA reductase and angiotensin-I converting enzyme and show antioxidative properties in endothelial EA. Hy 926 cells. *Food Science and Human Wellness*, 4(2), 80-85.
- 2) Ahn, J., Choi, W., Kim, S., & Ha, T. (2011). Anti-diabetic effect of watermelon (*Citrullus vulgaris* Schrad) on Streptozotocin-induced diabetic mice. *Food science and biotechnology*, 20(1), 251-254.
- 3) Ali, A. M., Gabbar, M. A., Abdel-Twab, S. M., Fahmy, E. M., Ebaid, H., Alhazza, I. M., & Ahmed, O. M. (2020). Antidiabetic Potency, Antioxidant Effects, and Mode of Actions of Citrus reticulata Fruit Peel Hydroethanolic Extract, Hesperidin, and Quercetin in Nicotinamide/Streptozotocin-Induced Wistar Diabetic Rats. *Oxidative medicine and cellular longevity*, 2020.

- 4) Ashraf, H., Butt, M. S., Iqbal, M. J., & Suleria, H. A. R. (2017). Citrus peel extract and powder attenuate hypercholesterolemia and hyperglycemia using rodent experimental modeling. *Asian Pacific journal of tropical biomedicine*, 7(10), 870-880.
- 5) Assefa, A. D., & Keum, Y. S. (2017). Effect of extraction solvent and various drying methods on polyphenol content and antioxidant activities of yuzu (*Citrus junos* Sieb ex Tanaka). *Journal of Food Measurement and Characterization*, 11(2), 576-585.
- 6) Avula, B., Sagi, S., Wang, Y.-H., Wang, M., Gafner, S., Manthey, J. A., & Khan, I. A. (2016). Liquid chromatography-electrospray ionization mass spectrometry analysis of limonoids and flavonoids in seeds of grapefruits, other citrus species, and dietary supplements. *Planta medica*, 82(11/12), 1058-1069.
- 7) Ben-Rejeb, I., Dhen, N., Kassebi, S., & Gargouri, M. (2020). Quality Evaluation and Functional Properties of Reduced Sugar Jellies Formulated from Citrus Fruits. *Journal of Chemistry*, 2020, 1-8. doi:10.1155/2020/5476872
- 8) Chang, Y. C., Chang, T. J., Jiang, Y. D., Kuo, S. S., Lee, K. C., Chiu, K. C., & Chuang, L. M. (2007). Association study of the genetic polymorphisms of the transcription factor 7-like 2 (TCF7L2) gene and type 2 diabetes in the Chinese population. *Diabetes*, 56(10), 2631-2637.
- 9) Chavan, P., Singh, A. K., & Kaur, G. (2018). Recent progress in the utilization of industrial waste and by-products of citrus fruits: A review. *Journal of Food Process Engineering*, 41(8), 1-10.
- 10) DeMambro, V. E., Kawai, M., Clemens, T. L., Fulzele, K., Maynard, J. A., De Evsikova, C. M., . . . Rosen, C. J. (2010). A novel spontaneous mutation of Irs1 in mice results in hyperinsulinemia, reduced growth, low bone mass and impaired adipogenesis. *Journal of Endocrinology*, 204(3), 241-253.
- 11) Dong, J., Cai, F., Shen, R., & Liu, Y. (2011). Hypoglycaemic effects and inhibitory effect on intestinal disaccharidases of oat beta-glucan in streptozotocin-induced diabetic mice. *Food chemistry*, 129(3), 1066-1071.
- 12) Estaji, M., Mohammadi-Moghaddam, T., Gholizade-Eshan, L., Firoozzare, A., & Hooshmand-Dalir, M.-A.-R. (2020). Physicochemical characteristics, sensory attributes, and antioxidant activity of marmalade prepared from black plum peel. *International Journal of Food Properties*, 23(1), 1979-1992.
- 13) Faid, S. M. A. E. F., Fadlalla, E. A. S., & Khojah, E. Y. (2017). Anti-diabetic and Antioxidant Effects of Grapefruit, Mango and Strawberry Juice in Streptozotocin-Induced Diabetic Rats. *Journal of Applied Life Sciences International*, 1-13.
- 14) FAO. (2016). *Processed*. Retrieved from
- 15) Farouk, H., El-Sayeh, B. A., Mahmoud, S. S., & Sharaf, O. A. (2012). Effect of olfactory stimulation with grapefruit oil and sibutramine in obese rats. *Journal of Pioneering Medical Sciences*, 2(1), 3-10.
- 16) Fayek, N. M., El-Shazly, A. H., Abdel-Monem, A. R., Moussa, M. Y., Abd-Elwahab, S. M., & El-Tanbouly, N. D. (2017). Comparative study of the hypocholesterolemic, antidiabetic effects of four agro-waste Citrus peels cultivars and their HPLC standardization. *Revista Brasileira de Farmacognosia*, 27(4), 488-494.
- 17) Gan, J., Peng, G., Liu, S., Hu, X., Wang, X., Guo, S., . . . Yu, Q. (2020). Comparison of structural, functional and in vitro digestion properties of bread incorporated with grapefruit peel soluble dietary fibers prepared by three microwave-assisted modifications. *Food and Function*, 11(7), 6458-6466.
- 18) Gerard, I.-R., Palomer, E., Ramos-Fernández, E., Guix, F. X., Bosch-Morató, M., Guivernau, B., . . . Ois, A. (2015). Fibrinogen nitrotyrosination after ischemic stroke impairs thrombolysis and promotes neuronal death. *Biochimica et Biophysica Acta Molecular Basis of Disease*, 1852(3), 421-428.

- 19) Huang, J.-y., Liao, J.-s., Qi, J.-r., Jiang, W.-x., & Yang, X.-q. (2020). Structural and physicochemical properties of pectin-rich dietary fiber prepared from citrus peel. *Food Hydrocolloids*, 110, 106140.
- 20) Hussein, A. M., Kamil, M., Hegazy, N., Mahmoud, K., & Ibrahim, M. (2015). Utilization of Some Fruits and Vegetables By-Products to Produce High Dietary Fiber Jam. *Food Science and Quality Management*, 37, 39-45.
- 21) Jiang, J., Yan, L., Shi, Z., Wang, L., Shan, L., & Efferth, T. (2019). Hepatoprotective and anti-inflammatory effects of total flavonoids of Qu Zhi Ke (peel of *Citrus changshan-huyou*) on non-alcoholic fatty liver disease in rats *via* modulation of NF- $\kappa$ B and MAPKs. *Phytomedicine*, 64, 1-9.
- 22) Kang, S., Song, S., Lee, J., Chang, H., & Lee, S. (2018). Clinical investigations of the effect of Citrus unshiu peel pellet on obesity and lipid profile. *Evidence-Based Complementary and Alternative Medicine*, 2018, 1-6.
- 23) Karbinger, M. S., Sierra, L., Minahk, C., Fonio, M. C., de Bruno, M. P., & Jerez, S. (2013). The role of oxidative stress in alterations of hematological parameters and inflammatory markers induced by early hypercholesterolemia. *Life sciences*, 93(15), 503-508.
- 24) Kesbiç, O. S., Acar, Ü., Yilmaz, S., & Aydin, Ö. D. (2020). Effects of bergamot (*Citrus bergamia*) peel oil-supplemented diets on growth performance, haematology and serum biochemical parameters of Nile tilapia (*Oreochromis niloticus*). *Fish Physiology and Biochemistry*, 46(1), 103-110.
- 25) Khalil, M. N., Farghal, H. H., & Farag, M. A. (2020). Outgoing and potential trends of composition, health benefits, juice production and waste management of the multi-faceted Grapefruit Citrus X paradisi: A comprehensive review for maximizing its value. *Critical Reviews in Food Science and Nutrition*, 1-22.
- 26) Kiefl, J., Kohlenberg, B., Hartmann, A., Obst, K., Paetz, S., Krammer, G., & Trautzsch, S. (2017). Investigation on key molecules of Huanglongbing (HLB)-induced orange juice off-flavor. *Journal of agricultural and food chemistry*, 66(10), 2370-2377.
- 27) Lawless, H. T., & Heymann, H. (2010). *Sensory evaluation of food: principles and practices*: Springer Science and Business Media.
- 28) Levent, N. B. K. A. H. (2014). Utilization of citrus albedo in tarhana production. *Journal of Food and Nutrition Research (ISSN 1336-8672)*, 53(2), 162-170.
- 29) Li, R., Zhang, Y., Rasool, S., Geetha, T., & Babu, J. R. (2019). Effects and underlying mechanisms of bioactive compounds on type 2 diabetes mellitus and Alzheimer's disease. *Oxidative medicine and cellular longevity*, 2019, 1-25.
- 30) Lin, L.-Y., Huang, C.-Y., Chen, K.-C., & Peng, R. Y. (2021). Pomelo fruit wastes are potentially valuable antioxidants, anti-inflammatories, antihypertensives, and antihyperglycemics. *Horticulture, Environment, and Biotechnology*, 62(3), 377-395.
- 31) Liu, Y., Heying, E., & Tanumihardjo, S. A. (2012). History, global distribution, and nutritional importance of citrus fruits. *Comprehensive reviews in Food Science and Food safety*, 11(6), 530-545.
- 32) Mahmoud, A. M. (2013). Hematological alterations in diabetic rats-role of adipocytokines and effect of citrus flavonoids. *Excli Journal*, 12, 647-657.
- 33) Marinelli, V., Lucera, A., Incoronato, A. L., Morcavallo, L., Del Nobile, M. A., & Conte, A. (2020). Strategies for fortified sustainable food: the case of watermelon-based candy. *Journal of food science and technology*, 1-8.
- 34) Maurya, A. K., Mohanty, S., Pal, A., Chanotiya, C. S., & Bawankule, D. U. (2018). The essential oil from Citrus limetta Risso peels alleviates skin inflammation: In-vitro and in-vivo study. *Journal of ethnopharmacology*, 212, 86-94.

- 35) Montgomery, D. C. (2017). *Design and analysis of experiments*: John Wiley and sons.
- 36) Nishad, J., Dutta, A., Saha, S., Rudra, S. G., Varghese, E., Sharma, R., . . . Kaur, C. (2020). Ultrasound-assisted development of stable grapefruit peel polyphenolic nano-emulsion: Optimization and application in improving oxidative stability of mustard oil. *Food Chemistry*, 334, 1-11.
- 37) Oboh, G., Olasehinde, T. A., & Ademosun, A. O. (2017). Inhibition of enzymes linked to type-2 diabetes and hypertension by essential oils from peels of orange and lemon. *International Journal of Food Properties*, 20(sup1), S586-S594.
- 38) Oyedemi, S., Adewusi, E., Aiyegoro, O., & Akinpelu, D. (2011). Antidiabetic and haematological effect of aqueous extract of stem bark of *Azelia africana* (Smith) on streptozotocin-induced diabetic Wistar rats. *Asian Pacific journal of tropical biomedicine*, 1(5), 353-358.
- 39) Oyetayo, F. L., Akomolafe, S. F., & Oladapo, I. F. (2019). A comparative study on the estimated glycemic index (eGI), phenolic constituents, antioxidative and potential antihyperglycemic effects of different parts of ripe *Citrus paradisi* fruit. *Oriental Pharmacy and Experimental Medicine*, 19(1), 81-89.
- 40) Özbek, T., Şahin-Yeşilçubuk, N., & Demirel, B. (2019). Quality and Nutritional Value of Functional Strawberry Marmalade Enriched with Chia Seed (*Salvia hispanica* L.). *Journal of Food Quality*, 2019, 1-8.
- 41) Pérez-Herrera, A., Martínez-Gutiérrez, G. A., León-Martínez, F. M., & Sánchez-Medina, M. A. (2020). The effect of the presence of seeds on the nutraceutical, sensory and rheological properties of *Physalis* spp. Fruits jam: A comparative analysis. *Food chemistry*, 302, 1-7.
- 42) Picot-Allain, M. C. N., Amiri-Rigi, A., Abdoun-Ouallouche, K., Aberkane, L., Djefal-Kerrar, A., Mahomoodally, M. F., & Emmambux, M. N. (2022). Assessing the bioactivity, cytotoxicity, and rheological properties of pectin recovered from citrus peels. *Food Bioscience*, 46, 1-9.
- 43) Plebani, M., & Piva, E. (2002). Erythrocyte sedimentation rate: use of fresh blood for quality control. *American Journal of Clinical Pathology*, 117(4), 621-626.
- 44) Putra, O., Saputro, I., Nurrahman, N., Herawati, E., & Dewi, L. (2020). Effects of empirical antibiotic administration on the level of C-Reactive protein and inflammatory markers in severe burn patients. *Annals of Burns and Fire Disasters*, 33(1), 20-26.
- 45) Sarfraz, M., Sajid, S., & Ashraf, M. A. (2016). Prevalence and pattern of dyslipidemia in hyperglycemic patients and its associated factors among Pakistani population. *Saudi Journal of Biological Sciences*, 23(6), 761-766.
- 46) Scurria, A., Sciortino, M., Albanese, L., Nuzzo, D., Zabini, F., Meneguzzo, F., . . . Avellone, G. (2021). Flavonoids in lemon and grapefruit IntegroPectin. *ChemistryOpen*, 10(10), 1055-1058.
- 47) Sir Elkhatim, K. A., Elagib, R. A., & Hassan, A. B. (2018). Content of phenolic compounds and vitamin C and antioxidant activity in wasted parts of Sudanese citrus fruits. *Food science and nutrition*, 6(5), 1214-1219.
- 48) Slemmer, J. E., Shaughnessy, K. S., Scanlan, A. P., Sweeney, M. I., & Gottschall-Pass, K. T. (2012). Choice of diet impacts the incidence of stroke-related symptoms in the spontaneously hypertensive stroke-prone rat model. *Canadian Journal of Physiology and Pharmacology*, 90(2), 243-248.
- 49) Soji-Omoniwa, O., Muhammad, N., Omoniwa, B., & Usman, L. (2014). Effect of Leaf Essential Oil of *Citrus sinensis* on Haematological Parameters of Alloxan-induced Diabetic Rats. *International Blood Research and Reviews*, 113-120.
- 50) Sreeramulu, D., Reddy, C., & Raghunath, M. (2009). Antioxidant activity of commonly consumed cereals, millets, pulses and legumes in India. *Indian J. Biochem. Biophys.*, 46(1), 112-115.

- 51) Tamer, C. E. (2012). A research on raspberry and blackberry marmalades produced from different cultivars. *Journal of Food Processing and Preservation*, 36(1), 74-80.
- 52) Teixeira, F., Santos, B. A. d., Nunes, G., Soares, J. M., Amaral, L. A. d., Souza, G. H. O. d., . . . Schwarz, K. (2020). Addition of Orange Peel in Orange Jam: Evaluation of Sensory, Physicochemical, and Nutritional Characteristics. *Molecules*, 25(7), 1670.
- 53) Tsujiyama, I., Mubassara, S., Aoshima, H., & Hossain, S. (2013). Anti-histamine release and anti-inflammatory activities of aqueous extracts of citrus fruits peels. *Oriental Pharmacy and Experimental Medicine*, 13(3), 175-180.
- 54) Udom, G. J., Yemitan, O. K., Umoh, E. E., Mbagwu, H., Ukpe, E. E., & Thomas, P. S. (2018). Hepatoprotective properties of ethanol seed extract of *Citrus paradisi* Macfad (Grape Fruit) against paracetamol-induced hepatotoxicity in wistar rats. *Journal of Herbal Drugs*, 8(4), 219-225.
- 55) Uzun, F. G., & Kalender, Y. (2013). Chlorpyrifos induced hepatotoxic and hematologic changes in rats: the role of quercetin and catechin. *Food and Chemical Toxicology*, 55, 549-556.
- 56) Vazquez-Olivo, G., Gutiérrez-Grijalva, E. P., & Heredia, J. B. (2019). Prebiotic compounds from agro-industrial by-products. *Journal of Food Biochemistry*, 43(6), 12711-12719.
- 57) Wang, B., Dong, T., Wang, J., Wang, H., Ma, M., Li, H., . . . Xiong, B. (2020). *Study on the accumulation of Phenols during fruit development of two Pomeles*. Paper presented at the IOP Conference Series: Earth and Environmental Science.
- 58) Wolf, B., & Weisbrode, S. (2003). Safety evaluation of an extract from *Salacia oblonga*. *Food and Chemical Toxicology*, 41(6), 867-874.
- 59) Younis, K., Islam, R. U., Jahan, K., Yousuf, B., & Ray, A. (2015). Effect of addition of mosambi (*Citrus limetta*) peel powder on textural and sensory properties of papaya jam. *Cogent Food and Agriculture*, 1(1), 1-8.
- 60) Zarate-Vilet, N., Gué, E., Delalonde, M., & Wisniewski, C. (2022). Valorization of Grapefruit (*Citrus paradisi*) Processing Wastes. In *Mediterranean Fruits Bio-wastes* (pp. 179-220): Springer.
- 61) Zhang, M., Zhu, J., Zhang, X., Zhao, D.-g., Ma, Y.-y., Li, D., . . . Huang, Q. (2020). Aged citrus peel (chenpi) extract causes dynamic alteration of colonic microbiota in high-fat diet induced obese mice. *Food and Function*, 11(3), 2667-2678.